INTRODUCTION

Diarrhea can occur due to food and beverage poisoning, with the highest cause being the result of infection with various bacteria, viruses, or parasites. Bacteria that can cause this disease are Escherichia coli bacteria, which are known as good bacteria in the digestive tract. But the reality is that in microbiology, not all types of Escherichia coli bacteria are good bacteria. Aim to find out the content and number of Escherichia coli bacteria colonies in UHT milk brands A and B as well as brand X yogurt products. Identification of Escherichia coli bacteria by the IMVIC Test method (Indole, Methyl-Red (MR) test, Voges Proskauer (VP), and Citrate). TPC (Total Plate Count), bacterial staining and microscope observation. Negative and positive results were obtained in the indole test and the methyl-red test was characterized by the formation of a red ring at the top for positive results and a yellow ring at the top for negative results, as well as negative results obtained in the Voges Proskauer test and the citrate test. Then for the results of gram staining and microscope testing the bacterial morphology was not seen. For the calculation of colonies, 45 colonies of sample A, 60 colonies of sample B, and 38 colonies of sample X. Samples containing Escherichia coli are contained in sample A of UHT milk and sample X of yogurt products and microbial contamination in samples according to SNI 2009.

Keywords: Escherichia coli, UHT Milk, Fermented Milk, IMVIC, TPC, yogurt

MATERIAL AND METHODS

Materials

The materials used in this study were UHT milk (samples A and B) and fermented milk (sample X) obtained from supermarkets in the Karawang area. Bacterial growth media are NA (Nutrient Agar), SIM (Sulfide-Indole-Motility), and MR-VP (Methyl Red-Voges Proskauer). The reagents used are Kovac reagents, methyl red reagents. Gram staining is the crystal violet, 96% alcohol, safranin, and aquadest.

Methods

Total Plate Count (TPC)

At Each Sample 1 mL is taken, then diluted using distilled water until dilution to 10⁶. Furthermore, From Each Sample 10⁶ Dilution, 1 mL is taken to be inserted into a sterile petri dish, then poured nutrient agar (NA) liquid media and homogenized by sliding horizontal plates or forming the number eight and allowed to freeze. incubation on milk 37°C for 24 hours and all colonies that grow are counted as TPC.

Biochemical Test

From the dilution tube, make a scratch on the nutrient agar (na) media, and incubate at 37°C for 18-24 hours. observe the murky white colonies of NA.

Indol Test

Colonies from na media were inoculated in a sim-filled tube and incubated at 35°C for 24 hours ± 2 hours. then add a few drops of the kovac reagent until a red ring appears on the top layer for positive results and yellow rings for negative results.

Methyl Red (MR) Test

Colonies from na media were inoculated in tubes containing 10 ml mr-vp and inoculated at 35°C for 48 hours ± 2 hours. then add 2-5 drops of the mr indicator to the tube. observe the presence of red for positive results and yellow for negative results.

Voges Prokauer Test (VP)

Pure bacterial isolate inoculated on mr-vp media and incubated for 24 hours at room temperature. Add 0.6 ml of 5% alpha naphtol solution followed by 0.2 mL of 40% KOH. observe the color change for 30 minutes.
Citrate Test

Inoculation of pure bacterial isolate by zig-zag scratch using ose or using inoculation needle on simmons citrate media to tilt, then incubate for 24 hours at room temperature. observe the color change from green to blue.

Gram Staining

In NA, take a murky white colony with ose, then place it on the glass preparation, fix it on the fire bypassing the glass preparation on the fire twice. drop gentian violet until the entire circle is covered, wait for 5 minutes. Clean over running water, then drop the lugol and wait for 1 minute. Clean again above running water. drop alcohol on the entire surface until no color wears off again. Clean again above running water. Drop safranin and wait for 2 minutes. clean again above running water. Dry preparations on a tissue.

Microscopic examination

The first drop of immersion oil, then check the preparations under the microscope of the smallest magnification first. After finding the colony lay, change the magnification up to 100 times. The appropriate form of escherichia coli is red, short trunk shape, and a single colony.

RESULTS AND DISCUSSION

Number of UHT Milk and Fermented Milk Colonies

Testing the number of colonies is done by using the cup calculation method with a 10^-6 dilution. In the calculation of the number of colonies test results obtained are shown in Figure 1 with calculations using the cup calculation method.

<table>
<thead>
<tr>
<th>Assays</th>
<th>Sample A</th>
<th>Sample B</th>
<th>Sample X</th>
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<tbody>
<tr>
<td>Indol</td>
<td>+</td>
<td>-</td>
<td>+</td>
</tr>
<tr>
<td>Methyl Red</td>
<td>+</td>
<td>-</td>
<td>+</td>
</tr>
<tr>
<td>Voges Prekauer</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Citrat</td>
<td>-</td>
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Table 1 Biochemical Test Results

From the data above sample X which is a sample of fermented milk has the lowest number of colonies compared to samples A and B which are UHT milk samples. This can happen because the level of dilution is carried out low so that the growth of microorganisms becomes very numerous and difficult to do calculations, whereas for samples A and B the level of dilution is carried out high (Caplan & Barbano, 2013). For samples A and B which are flavorful and non-flavored UHT milk, the results of the calculation of the number of colonies are in accordance with SNI provisions in 2009 concerning the maximum limit of microbial contamination, namely for flavored and non-flavored UHT milk <10^5 colonies / 0.1 mL or 100 colonies / 1 mL, while for sample X which is fermented milk is also in accordance with the provisions of SNI in 2009 concerning the maximum limit of microbial contamination that is max 10^5 / 0.1 mL or 100 colonies / 1 mL.

Identification of Escherichia coli bacteria

Tests in biochemical tests include various tests to determine the activity of microbial metabolism. Observation of metabolic activity is known to be the ability of microbes to use and decompose complex molecules such as starches, fats, proteins, and nucleic acids.

The Indol Test

The Indole test aims to identify the ability of bacteria to produce indole by using the enzyme tryptophanase. Indole production in the media is possible because of tryptophan. Tryptophan is an essential amino acid, which is oxidized by several bacteria which results in the formation of indole, pyruvic acid, and ammonia.

The Methyl Red (MR) Test

The Methyl Red (MR) Test aims to detect the ability of organisms to produce and maintain the final acid-stable product from glucose fermentation. Methyl red is a pH indicator, which remains red at pH 4.4 or less. The observations for the MR test yielded positive results as indicated by the presence of a red ring on the top.

The Voges Proskauer (VP) test

The Voges Proskauer (VP) test is a test used to detect acetonin in a liquid culture of bacteria. This test is done by adding alpha-naphthol and potassium hydroxide. The red color indicates a positive result, while the yellow-brown or colorless color is a negative result. This test is negative for Escherichia coli because Escherichia coli ferments carbohydrates into acidic products and does not produce neutral products such as acetonin.
The citrate test aims to detect the ability of an organism to utilize it as the only source of carbon and energy. If the bacteria are able to use citrate as their carbon source, it will increase the pH and change the color of the culture medium from green to blue.

Observation results for the citrate test yielded negative results indicated by the absence of color changes in the citrate test media. Biochemical tests are carried out to determine the physiological properties of bacteria. Based on biochemical tests, the results showed that *Escherichia coli* bacteria were found in samples A and X. Positive tests on samples A and X were marked by the formation of a red ring on the upper layer of the indole test and the methyl red test. For negative results, sample B shows the results of non-*Escherichia coli* bacteria or other coliform bacteria, this negative result is also marked by the formation of yellow rings in the upper layer of the indole test and methyl red test. However, the results of this biochemical test have not shown specific results. Therefore the next test is gram staining and observation of the sample under a microscope.

UHT milk samples that produce positive results indicate incompatibility with the provisions or unfavorable research treatments such as samples contaminated with bacteria from outside during processing (Boor et al., 2017). The test results on UHT milk samples indicate the presence of *Escherichia coli* bacteria is not in accordance with the provisions or unfavorable research treatments such as samples contaminated with bacteria from outside during processing (Boor et al., 2017). The test results on UHT milk samples indicate the presence of *Escherichia coli* bacteria which results in no microbial contamination in UHT milk. Whereas UHT milk in sample B produced negative results showing conformity with the conditions specified for the process of making UHT milk. These results indicate that the manufacturing process in sample B follows the applicable terms or conditions (Coppa et al., 2013). So the product produced is good because UHT milk is milk with a manufacturing process using heating techniques at high temperatures allowing the bacteria contained in the milk to die. For yogurt (sample X) that produces positive results containing *Escherichia coli* bacteria is a sample of fermented milk (Aryana & Olson, 2017). This happens because the possibility of fresh milk used in the manufacture of fermented milk has been contaminated with *Escherichia coli* bacteria. So the product produced contains *Escherichia coli* bacteria (Butler et al., 2011). Because fermented milk is milk that is inoculated with lactic acid bacteria, the results of the methyl red test produce positive results because the lactic acid bacteria itself is able to ferment sugar into lactic acid (Boor et al., 2017). Another cause of the presence of *Escherichia coli* bacteria in the sample is the possibility of antibacterial activity against undesirable bacteria such as *Escherichia coli* (Martin et al., 2011). This can happen because the lactic acid production process runs fast so that the growth of other microbes that are not used in mixing can be inhibited like *Escherichia coli*. This statement is not necessarily true whether the bacteria contained in the sample used are *Escherichia coli* bacteria or other bacteria. For this reason, antibacterial activity testing must be done on the sample (Kouskhi et al., 2016).

**Gram Staining and Microscope Test**

Gram staining is used to identify bacteria. Bacteria stained by the gram method are divided into two groups, namely gram-positive bacteria and gram-negative bacteria. Gram-positive bacteria are bacteria that do not maintain purple dyes in gram staining. While gram-positive bacteria are bacteria that will maintain a dark purple color after rinsing with alcohol (Tjatur et al., 2015).

The results of gram staining and testing under a microscope showed that the morphology did not appear to be due to the process of making preparations that were too thick (Figure 10), making preparations that were too thin (Figure 9), preparations that were not completely dry, bacteria that did not carry over from bacterial culture at the time of collection using an ose needle and improper treatment. It cannot provide information about the color of bacterial cells or show the nature of gram bacteria and cell shape. Errors in making preparations have also been explained by (Widyastuti et al., 2014) that things that often fail microscopy testing is excessive culture taking, preparations that are too thick, preparations that are too thin, preparations that have not been completely dry resulting in microorganisms which were observed to be poorly formed, as well as a less aseptic treatment aimed at avoiding contamination. In these results did not show specific results, then conducted a comparison test with samples inoculated with *Escherichia coli* bacteria (Butler et al., 2011).

**Figure 8** The Voges Proskauer test sample A, B dan X (-)

**Figure 9** The citrate test sample A, B dan X (-)

**Figure 10** Preparations that are too thick (sample B and X)

**Figure 11** Preparations that are too thick (sample B and X)
Escherichia coli O157:H7 SURVIVAL IN TRADITIONAL AND LOW PASTEURIZATION TEMPERATURES.


